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Animal health and greenhouse gas intensity: the paradox of periparturient parasitism

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1 **Animal health and greenhouse gas intensity: the paradox of**
2 **periparturient parasitism**

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Abstract

Here we provide the first direct measurements of pathogen challenge impacts on greenhouse gas (GHG) production, yield and intensity. Twin-rearing ewes were *ad libitum* fed pelleted lucerne from day₋₃₂ to day₃₆ (day₀ is parturition), and repeatedly infected with 10,000 *Teladorsagia circumcincta* infective larvae (n=16), or sham-dosed with water (n=16). A third group of 16 ewes were fed at 80% of uninfected ewes' feed intake during lactation. Methane emissions were measured in respiration chambers (day₃₀ to day₃₆) whilst total tract apparent nutrient digestibility around day₂₈ informed calculated manure methane and nitrous oxide emissions estimates. Periparturient parasitism reduced feed intake (-9%) and litter weight gain (-7%) and doubled maternal body weight loss. Parasitism reduced daily enteric methane production by 10%, did not affect methane yield per unit dry matter intake but increased yield per unit digestible organic matter intake by 14%. Parasitism did not affect daily calculated manure methane and nitrous oxide production, but increased manure methane and nitrous oxide yields per unit dry matter intake by 16% and 4%, respectively, and per unit digestible organic matter intake by 46% and 31%, respectively. Accounting for increased lucerne input for delayed weaning and maternal body weight loss compensation, parasitism increased calculated GHG intensity per kg lamb weight gain for enteric methane (+11%), manure methane (+32%) and nitrous oxide (+30%). Supplemented with the global warming potential (GWP) associated with production of pelleted lucerne, we demonstrated that parasitism increased calculated GWP per kg lamb weight gain by 16%, which was similar to the measured impact of parasitism on feed conversion ratio. Thus, arising from

40 pathogen-induced feed efficiency reduction and modified GHG emissions, we
41 demonstrated that ovine periparturient parasitism increases GHG intensity.
42 This implies that ewe worm control can not only improve production efficiency
43 but also reduce the environmental footprint of sheep production systems.

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45 **Key words:** disease, parasitism, environmental footprint, methane, nitrous
46 oxide, sheep

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1. Introduction

It is well recognized that pathogen exposure often results in anorexia, i.e. reduction in feed intake. In the case of sub-clinical gastrointestinal nematode parasitism, feed intake is typically reduced by up to 20-25% in e.g. growing and periparturient sheep, though wide ranges of parasitism-induced anorexia and associated production losses have been reported across different species (Sykes, 1994; Kyriazakis et al., 1998; Zaralis et al., 2008). Variation in feed intake can be expected to correlate with variation in greenhouse gas (GHG) production from both respiration and manure emission. This implies that pathogen challenge would be expected to result in reduced daily production of methane (CH_4), carbon dioxide (CO_2) and nitrous oxide (N_2O), provided that GHG yield, defined as the amount of GHG produced per unit feed intake, is not affected. However, since pathogen challenge reduces productivity, arising from a combination of anorexia and reduced efficiency of resource use for production purposes (Sykes, 1994; Coop and Kyriazakis, 1999), challenged animals would be expected to take longer and require more resource input to achieve the same productive output. GHG production associated with this extra resource input required would effectively be the consequence of pathogen challenge on resource efficiency, and thus increase GHG intensity.

Here, we provide the first direct assessment of the impact of pathogen challenge on GHG emission in livestock. We have assessed effects of gastrointestinal parasitism on performance, digestibility, CH_4 and CO_2 production and yield, and on feed efficiency, in lactating ewes. Furthermore, we used IPCC (2006) assumptions and literature data where data were not

derived from the experiment carried out to extend the above to include estimates of manure CH₄ and N₂O production and yield. These estimates are used to test the hypothesis that periparturient parasitism increases ewe GHG intensity and global warming potential (GWP) for lamb production.

2. Materials and Methods

2.1 Animals and housing

Twelve 4-5 year old Mule ewes (Bluefaced Leicester × Scottish Blackface) were recruited from each of four larger mating group approximately 45 days before observed mean parturition date (day₀), with mean expected parturition dates separated by a week. Ewe body weight (BW) and body condition score (CS) recorded on day₋₃₉ for the total of 48 ewes used averaged (±se) 68.2±0.79 kg and 2.5±0.06, respectively. Ewes were served by Suffolk rams and confirmed to be bearing twins by ultrasonic scanning prior to the experiment and were housed individually, in pens sized 1.30 m × 2.15 m with adjacent creep area of the same size for their lambs. From day₋₄₅ until day₃₀, the ewes were housed in a naturally ventilated and illuminated shed, with additional low-level lighting at night times during lambing. From day₃₀ until day₃₆, ewes and their lambs were housed in respiration chambers (see below) in similar sized pens. Fresh wood shavings were used as bedding and added daily, and fresh water was *ad libitum* available. A small amount of shavings were also used daily in the respiration chambers.

2.2. Experimental treatments and feeding

The twelve ewes within each of the four mating groups were divided into three groups of four ewes based on initial BW, which resulted in three groups of 16 ewes with similar mean initial BW, CS and faecal egg counts (FEC). From day₋₄₅ to day₋₃₂, the ewes received *ad libitum* medium quality hay and approximately 300 g/day/head of a commercial ewe nut. From day₋₃₂ until day₋₂₅, allowances of hay and commercial ewe nuts were gradually reduced and completely replaced with increasing amounts of pelleted lucerne. From day₋₂₄ onwards, two groups of ewes were fed lucerne *ad libitum* and either uninfected (CON) or dosed with parasites (PAR). Details of the experimental infection are provided below. A third group of ewes were managed as CON ewes during pregnancy but fed restrictedly at 80% of intakes achieved by CON ewes during lactation (RES). The RES group was included to assess to what extent GHG production, yield and intensity would be affected by reduced feed intake per se. Ewes were fed at 07.30 and 1500 h. The experiment was approved by SRUC's Ethical Review Committee (ED AE 03/2011) and carried under Home Office authorization (PPL 60/3782).

2.3. Experimental infection

Because the ewes were 4 to 5 years old and had previously grazed natural pastures infested predominantly with *Teladorsagia circumcincta*, they were expected to have had substantial prior exposure to this parasite, an abomasal nematode of particular concern in temperate regions. The ewes were orally treated on day₋₃₈ with levamisole (Levacide, Norbrook, Newry, UK) and ivermectin (Oramec, Merial, Harlow, UK) at the rate of 7.5 and 0.2 mg/kg BW, respectively, to remove worm burdens. A subsequent FEC taken on day.

22 averaged 0 (0-1) eggs per g fresh faeces (epg), suggesting that the drench was effective. The PAR ewes were then trickle infected with 10,000 infective *T. circumcincta*, suspended in 10 ml of water and administered every Monday, Wednesday and Friday from day-21 onwards until the end of the experiment. The CON and RES ewes were sham-infected with 10 ml of water on the same days. The *T. circumcincta* strain used was the Moredun Ovine Susceptible Isolate that has been maintained in the laboratory for several years. This infection model has repeatedly been used in our lab to induce sub-clinical parasitism in periparturient ewes (Houdijk et al., 2003, 2006, Zaralis et al., 2009; Kidane et al., 2010).

2.4. Measurements and calculations

Performance. The ewes were weighed on day-39 and then weekly from day-31 onwards, as well as within 12 h of parturition to assess daily weight gain during late pregnancy and during lactation through linear regression of BW on time. The lambs were weighed within 12 h after birth and weekly afterwards to assess daily litter weight gain in the same way. Since the lambs did not receive creep feed, lamb BW and daily weight gain were used to calculate milk production (Robinson et al., 1969). Ewe CS was taken approximately fortnightly, by lumbar palpation on a zero to five point scale, and to an accuracy of a quarter (Russel et al., 1969), where 0 is emaciated and 5 is obese. Feed samples were collected every day during the experiment during feeding and were pooled for chemical analyses (Table 1) as per standard protocols (Ministry of Agriculture Fisheries and Food, 1992). Feed refusals were recorded twice weekly (Mon and Thu) and analysed for dry

matter (DM) only. This allowed for the calculation of achieved mean daily dry matter intake (DMI).

Parasitism. The level of parasitism was monitored through regular faecal sampling for FEC, according to a modified flotation method (Christie and Jackson, 1982), with a sensitivity of 1 epg. This was done for all ewes at housing, day-22 and day-11 and at parturition, and then weekly thereafter (for PAR ewes only).

Digestibility. Apparent total tract DM, organic matter (OM) and nitrogen (N) digestibility were assessed through using acid insoluble ash (AIA) as an internal, indigestible marker. Feed samples collected daily during feeding were pooled for DM, N, ash and AIA analyses, with OM calculated as DM minus ash. Faeces were collected directly from the rectum of all ewes for three consecutive days (day₂₇ to day₂₉) and were pooled per individual ewe and kept frozen at -20 °C before analysis of DM, ash, AIA and N, and calculation of OM as above. Feed and faecal AIA were analysed using the 2 M HCl procedure described by van Keulen and Young (1977). The above analyses allowed us to calculate faecal OM and N output, as well as digestible OM intake (dOMI).

Furthermore, daily fresh faeces production was calculated using mean achieved DM intake, faeces DM contents and total tract DM digestibility in order to estimate worm egg output (eggs/day). The latter was estimated by multiplying mean FEC during lactation (eggs/g) with the calculated fresh faeces production (g/day), under the assumption that since ewes were fed the same diet throughout lactation, total tract DM digestibility measured from

day₂₃ to day₂₆ can be extrapolated over the whole lactation period (Kidane et al. 2009).

Enteric methane and carbon dioxide emissions. Staggered lambing arising from the four mating groups used allowed for four rounds of housing in one of six indirect open-circuit respiration chambers (No Pollution Industrial Systems Ltd., Edinburgh, UK) for six days from day₃₀ to day₃₆ with two ewes per treatment per chamber, individually housed with their lambs to achieve two treatment replicates per round, and thus n=8 in total. Experimental treatments and DMI determination were maintained in chamber. Daily CH₄ and CO₂ production was measured as described in detail elsewhere (Rooke et al., 2014). Briefly, each chamber has an area of 25.4 m² with appropriate penning for two ewes and their four lambs. Air was removed from the chambers by exhaust fans set at 50 litre/s and temperature and humidity were set at 15 ± 1°C and 60 ± 5% relative humidity, respectively. Exhaust air was sampled for gas analysis sequentially for 45 s from each chamber, and methane and carbon dioxide concentrations were measured by infrared absorption spectroscopy. Animals remained in the chambers for 6 days, where the first four days were used for adaptation, and the last two days were used to quantify CH₄ and CO₂ production. Measurements of CH₄ and CO₂ concentrations were made every 6 minutes in the mechanically ventilated air entering and leaving each chamber and exhaust air flow rate (every 30 min) corrected to standard temperature and pressure. Due to missing data arising from operational issues in Round 2, a full data set for each room was available for only the last 24 h, which was the dataset used for the present

analysis. Daily CH₄ and CO₂ production was divided by mean daily DMI and dOMI to obtain CH₄ and CO₂ yield.

Manure methane emissions. Methane emissions from manure were estimated from the volume of volatile solids produced, defined as the sum of faecal and urine OM output (IPCC, 2006). Faecal OM output was derived from mean daily in chamber DMI, feed OM content and total tract OM digestibility, whilst total urine OM output was calculated under the assumption that energy excretion in urine is $0.04 \times \text{GE intake}$ (IPCC, 2006) and that energy concentration in urine is 18.75 MJ/kg DM (IPCC, 2006). We further assumed that the manure is directly deposited on pasture in a cool climate (i.e. not stored), and thus a methane conversion factor of 1% and a maximum methane-producing capacity of 0.19 m³ per kg volatile solids (IPCC, 2006). This resulted in a methane yield of 1.273 g per kg volatile solids. Daily manure CH₄ production was divided by mean daily in chamber DMI and dOMI to obtain manure CH₄ yield estimates.

Manure N₂O emissions. The N₂O emission from manure was estimated from total N excreted, i.e. the sum of faecal N and urine N outputs. Faecal N output is derived from mean daily in chamber DMI, feed N content, and apparent total tract N digestibility. Urine N output is estimated from the assumption that urine N output equals $0.46 \times \text{N apparently absorbed during lactation}$. This coefficient was derived from a series of then N balance estimates in lactating ewes (Lynch et al., 1988; Malik et al., 1999; Pappas, 1977; Maamouri et al., 2011), whilst N absorbed was derived from apparent total tract N digestibility carried out. As above, we assume that the manure is deposited directly on the pasture in a cool environment, and 1% of manure N

is directly volatilised into nitrous oxide N, 20% of manure N is indirectly volatilised through ammonia with an efficiency factor of 0.01 for conversion into nitrous oxide N and 30% of manure N is leached through nitrate with an efficiency factor of 0.0075 for conversion of manure N into nitrous oxide N (IPCC, 2006). This resulted in a N₂O yield estimate of 22.393 g per kg N excreted. Resulting daily N₂O production was divided by mean daily DMI and dOMI to obtain N₂O yield.

Greenhouse gas emission intensity calculations. The GHG yields derived as above were used to calculate GHG emission intensity per functional unit, which was one kg lamb BW gain (BWG) from parturition until weaning. We used the observed DMI, ewe BW loss, lamb birth weight and lamb BW gain until day₃₆, to calculate through extrapolation the number of days and the amount of DMI needed, as well as the total ewe BW loss incurred, to reach a target weaning weight of 25 kg live weight for each of two lambs reared (Kidane et al., 2010). This DMI was multiplied with GHG yields as measured for CON, PAR and RES ewes.

We also calculated DMI needed to restore final ewe BW to initial ewe BW (day-39), to account for BW loss incurred as a resource used to rear lambs to weaning. We assumed that the metabolizable energy (ME) requirement for restoring ewe BW was 39.75 MJ/kg for CON and RES ewes (AFRC, 1993) but 49.16 MJ/kg for PAR ewes for reasons outlined below (see 'sensitivity analysis'). Since any influence of PAR or RES treatment on digestibility and efficiency of resource utilization for weight gain post weaning would unlikely remain present post weaning, as ewes return to full immunity to parasites when lactation ceases (Houdijk, 2008), DMI to restore ewe BW was

multiplied with mean GHG yields for CON ewes. Total DMI was divided over total lamb weight gain to calculate feed conversion ratio. Furthermore, post weaning urine N output was estimated from the assumption that urine N output equals $0.76 \times \text{N apparently absorbed}$; this coefficient was derived from a series of then N balance estimates in growing sheep (Akinbamijo et al., 1994; Lima et al., 2011; Gorniak et al., 2014; Kimambo et al., 1988).

GHG emission intensity data from the above described CH_4 and N_2O sources were combined into one GWP figure after converting CH_4 and N_2O into their CO_2 equivalents. Conversion factors used were 25 and 298 $\text{CO}_2\text{-eq}$ for methane and nitrous oxide, respectively (IPCC, 2006). Data from CH_4 and N_2O were supplemented with a GWP of 320 g $\text{CO}_2\text{-eq/kg}$ for the production, dehydration and transport of lucerne with 11% residual moisture (Gallego et al., 2011), to derive at total GWP ($\text{kg CO}_2\text{-eq/kg BWG}$). Thus, breath CO_2 was not included in these calculations, as GWP assessments focus on net contributors to global warming, i.e. fossil-derived CO_2 only. We also used lamb BWG to estimate milk production (Robinson et al., 1969), in order to estimate GHG intensity per kg milk.

Sensitivity analysis. It was deemed appropriate to use a greater requirement of ME per kg BW gain to restore BW for PAR ewes than for CON and RES ewes, as PAR ewes likely lost relatively more fat than did CON and RES ewes (see results on CS). The 49.16 MJ/kg BW gain was arbitrarily chosen as the average of 39.75 MJ/kg (AFRC, 1993) and 58.57 MJ/kg (Olthoff et al. 1989), which would be requirements for fat deposition only. A sensitivity analysis was undertaken for the impact of this assumption by

comparing GWP intensity for PAR ewes to that of CON and RES ewes for ME requirements per kg BW gain taken as 39.75, 49.16 and 58.57 MJ/kg.

Since there are no data on the effect of parasitism in general, and of *T. circumcincta* challenge in particular, on urine N excretion in lactating sheep, we assumed that the aforementioned urine N as a proportion of apparent N absorbed is similar between CON, PAR and RES ewes. However, earlier studies showed that *T. circumcincta* infections in young lambs increased urinary N output as a proportion of apparent N absorbed by 30 to 40% relative to non-infected control lambs (Parkins et al., 1973; Sykes and Coop, 1977). We therefore undertook a second sensitivity analysis by comparing GWP intensity for urine N output by increasing the value of $0.46 \times \text{N absorbed}$ to 0.65 (40% increase) during lactation.

2.5. Statistical analysis

Data obtained during late pregnancy and early lactation were analysed separately, whilst the effect of parasitism during late pregnancy was analysed through comparing PAR ewes with combined CON/RES ewes. During late pregnancy, parameters that were repeatedly taken, i.e. feed intake, ewe BW and CS were analysed via repeated measures ANOVA. Parameters that were measured once, i.e. ewe and litter BW at birth, were analysed through ANOVA. Likewise, during lactation, ewe feed intake, BW, CS and hourly methane and carbon dioxide production were analysed via repeated measures ANOVA, whilst ewe and litter BW gain, digestibility figures, daily GHG production, yield and intensity data were analysed through ANOVA.

Models included mating group as block and day₋₃₉ observations as covariate where appropriate. The ewe was the experimental unit for production, parasitological and digestibility observations (n=16). However, for any parameter related to emission production, yields and intensities, the respiration chamber with paired ewes was used as experimental unit (n=8). Means were separated through the use of Fisher's protected LSD test at $P < 0.05$.

3. Results

3.1. Performance during late pregnancy

Figure 1 shows mean food intake over time during both late pregnancy and early lactation. Treatment did not interact with time for DM intake during late pregnancy ($P=0.33$), which averaged 3.02 and 2.77 kg/day for the combined CON/RES ewes and PAR ewes, respectively (s.e.d. 0.13 kg/day; $P=0.05$). Time and treatment did not interact for BW during late pregnancy ($P=0.70$), reflected in an numerically greater BW gain for the combined CON/RES ewes than for PAR ewes at 272 and 228 g/day, respectively (s.e.d. 31 g/day, $P=0.11$). CON/RES ewes tended to be heavier at parturition than PAR ewes, averaging 70.1 and 68.0 kg, respectively (s.e.d. 1.19 kg; $P=0.08$), though litter BW at birth did not differ, averaging at 9.07 and 9.30 kg, respectively (s.e.d. 0.44 kg; $P=0.61$). Treatment did not interact with time for CS ($P=0.47$) during late pregnancy, which gradually increased from 2.51 ± 0.06 on day₋₃₉ to 2.72 ± 0.06 on day₋₆ ($P=0.017$).

3.2. Total tract apparent digestibility and performance during early lactation

Table 2 shows that the total tract DM, OM and CP digestibilities were smaller in PAR ewes than in both CON and RES ewes ($P<0.05$). As for late pregnancy, feed intake and time did not interact during early lactation (Figure 1; $P=0.33$). Table 2 shows that mean DM intake was greater for CON ewes than for PAR ewes, which was in turn greater than for RES ewes. However, PAR and RES ewes achieved similar levels of dOM and digestible CP intake, which were both smaller than those for the CON ewes. Ewe BW loss was less for CON ewes than for PAR ewes, which in turn was less than for RES ewes. However, litter BW gain and calculated milk production were greater for CON ewes than for PAR and for RES ewes, whilst both did not differ between PAR and RES ewes (Table 2).

Treatment tended to interact with time for CS ($P=0.07$) during lactation; CS averaged 2.46 ± 0.06 across treatments on day₆ and 2.08, 1.84 and 2.02 on day₂₉ for CON, PAR and RES, respectively (se 0.07), suggesting that CS reduced for all ewes ($P<0.01$) but at a higher rate for PAR from ewes than for CON and RES ewes during lactation.

3.3. Parasitism

Ewe FEC averaged 101 (86 - 119) epg on day₋₃₉ and 0 (0 - 1) epg at day₋₂₂. Following infection from day₋₂₁ onwards, FEC of CON/RES and PAR ewes averaged 1 (1-2) and 0 (0-1) epg on day₋₁₁, respectively, and 3 (2-4) and 26 (15-43) epg at parturition, respectively ($P<0.001$). The FEC of PAR ewes then gradually increased to 71 (48 to 106) epg by day₂₈. Combined with

calculated fresh faeces production, the latter translated into 962 (675-1371) x 10³ worm eggs per day.

3.4. GHG production and yield

Figure 2 displays the average hourly CH₄ (Fig 2a) and CO₂ (Fig 2b) production of sets of two CON, PAR and RES ewes. Experimental treatment interacted with time for both GHG (P<0.001), arising from the larger diurnal variation in the RES ewes. Table 3 shows enteric and manure GHG production and yield per kg dry matter intake (DMI) as well as per kg digestible organic matter intake (dOMI), expressed both as CH₄ and N₂O and converted to CO₂ equivalents. We observed that CON ewes produced more enteric CH₄ than PAR ewes, which in turn produced more enteric CH₄ than RES ewes. However, whilst enteric CH₄ yield per kg DMI did not differ, averaging 10.23 g per kg DMI, enteric CH₄ yield per kg dOMI was ~14% greater for PAR ewes than for CON and RES ewes (P<0.05). We observed similar patterns for CO₂ emissions, though CO₂ production did not differ between CON and PAR ewes (P=0.15), CO₂ yield per kg DMI did not differ, averaging 690 g, whilst CO₂ yield per kg dOMI for RES ewes was intermediate to that of CON and PAR ewes, whose CO₂ yields differed (P<0.05).

The pattern in manure GHG emissions differed from enteric GHG emissions. CON and PAR ewes produced similar volumes of manure volatile solids, and thus CH₄, per day though manure CH₄ yield was greater for PAR ewes than for CON ewes, both per kg DMI (P<0.05) and per kg dOMI (P<0.01). Likewise, CON and PAR ewes produced similar amount of manure

N, and thus N₂O, per day though N₂O yields were greater for PAR ewes than for CON ewes, both per kg DMI (P=0.082) and per kg dOMI (P<0.05).

3.5. GHG intensity and sensitivity analysis

Table 4 shows the outcomes of the underlying calculations towards the treatment effects on GHG intensity. PAR and RES ewes required on averaged ~5 days longer than CON ewes to reach the target lamb weaning BW. This was associated with the same total DMI for CON and PAR ewes, but a significantly smaller DMI for RES ewes (P<0.01). Total BW loss was greater for PAR ewes than for CON ewes (P<0.05), and in turn greater for RES ewes than for PAR ewes (P<0.05). The estimated amount of DMI needed to restore BW lost was smaller in CON ewes than in PAR and RES ewes (P<0.01). However, the two pools of DMI combined were very similar for CON and RES ewes but greater for PAR ewes (P<0.05). The latter was reflected in a greater feed conversion ratio (P<0.05). Manure volatile solids and N output were greater for PAR ewes than for CON and RES ewes.

Table 4 also shows that the resulting GHG intensity for each of the underlying sources was greater for PAR ewes than for CON and RES ewes. Combined with the GWP for lucerne production, our calculations show that the GWPs for lamb production of CON, PAR and RES ewes were 5.09, 5.91 and 5.28 kg CO₂-eq per kg lamb weight gain, respectively, which was calculated to correspond with 1.04, 1.24 or 1.10 kg CO₂-eq per kg milk, respectively (s.e.d. 0.04 kg CO₂-eq per kg; P<0.05).

The sensitivity analysis indicated that GWP per kg lamb BWG was not very sensitive to variation in urine N excretion; an increase in urine N from

0.46 to 0.60 × N absorbed during lactation increased GWP by less than 1.4% from 5.91 to 5.99 CO₂-eq per kg lamb weight gain. However, GWP was more sensitive to variation in ME requirement for ewe BW gain; GWP for PAR ewes varied from 5.69 to 5.91 to 6.13 kg CO₂-eq/kg BWG for ME requirements per kg BW gain taken as 39.75, 49.16 and 58.57 MJ/kg, respectively. Nevertheless, each of these figures were significantly greater than the 5.09 kg CO₂-eq/kg BWG for the CON ewes.

4. Discussion

Here we propose a framework that accounts for pathogen-induced variation in GHG yield and reduction in feed efficiency in order to experimentally test the hypothesis that periparturient parasitism increases ewe GHG intensity for lamb production. To our knowledge, this is the first set of data that empirically addresses the consequences of impaired animal health on GHG intensity. The data obtained support the view that whilst ovine periparturient gastrointestinal nematode parasitism reduced GHG production per day, it paradoxically resulted in an increased GHG intensity for their lamb BWG by ~16%. Given that the latter was of similar magnitude to the impact of ewe parasitism on feed conversion ratio, the calculated increased GHG intensity largely came from accounting for the impact on production losses rather than on GHG yield per kg dry matter intake (DMI). As such, impact of animal health on GHG intensity is driven by a combination of a reduction in feed intake and feed nutritive value, the latter illustrated by ewe parasitism increasing GHG yield per kg digestible organic matter intake (dOMI).

417 Periparturient parasitism evoked anorexia in our ewes, especially
418 during lactation, but also reduced milk production and accelerated ewe BW
419 loss, which is in support of earlier observations (Leyva et al., 1982; Zaralis et
420 al., 2009). Although the degree of anorexia was relatively small at 9%, its
421 effect on litter weight gain (milk production) and ewe BW loss would have
422 been exacerbated by the negative impact of parasite challenge on total tract
423 nutrient digestibility, which agrees with other studies (Parkins et al., 1973;
424 Sykes and Coop, 1977). The impact on OM digestibility, which can be seen as
425 a proxy for energy digestibility, resulted in a reduction in dOMI of ~26% for
426 PAR ewes relative to CON ewes. The PAR ewes minimised the impact of the
427 latter on ME availability by mobilising more body fat, as the rate and extent of
428 body condition score loss tended to be greater than in CON ewes.
429 Consequently, it was assumed that the ME requirements per kg BW gain for
430 replenishment of body reserves for PAR ewes upon weaning was also greater
431 than that for CON ewes, due to the higher energy contents of fat (AFRC,
432 1993). The sensitivity analysis for the arbitrarily taken ME requirement for BW
433 gain of 49.16 MJ/kg as the average of 39.75 MJ/kg for average body
434 composition (AFRC (1993), and 58.57 MJ/kg for fat deposition only (Olthoff et
435 al., 1989), indicated that the impact of ovine parasitism on calculated GHG
436 intensity for lamb weight gain would reduce to 12% if standard assumptions
437 are taken but remains significant, with the largest contribution coming from
438 N₂O. Calculated GHG intensity for lamb weight gain was not very sensitive to
439 urinary N excretion; increasing the latter by up to 40%, i.e. the effect of
440 parasitism on N balance in growing lambs (Parkins et al., 1973; Sykes and
441 Coop, 1977), increased GHG intensity for lamb weight gain by less than 2%.

This is reflective of the relatively small proportion of nitrogen intake that is excreted with the urine in lactating ewes, where a significant proportion of dietary nitrogen is excreted with the milk.

Compared to our previous studies where similar infection protocols were used in periparturient ewes (Houdijk et al., 2003, Houdijk et al., 2006; Kidane et al., 2010), FEC of the PAR ewes were relatively low with levels below 100 epg. In addition, achieved DMI was relatively high, typically ~60% greater than that of ewes that were fed at 90% of assumed ME requirements (Kidane et al., 2010). The low FEC observed may have been the outcome of concurring high intake levels of metabolizable protein (MP), which is known to reduce ewe FEC through improved host resistance to nematodes (Houdijk et al., 2012). Since *in vitro* OM digestibility (Table 1) was very similar to *in vivo* OM digestibility (Table 2) for CON ewes, we can assume that the reduction in total tract OM and CP digestibility (Table 2) for PAR ewes proportionally reduced book values of fermentable ME and by-pass protein content (Hazzledine, 2014). Consequently, consumed lucerne pellets may have yielded ~70 g MP per kg DMI for PAR ewes, compared to ~80 g MP per kg DMI in CON and RES ewes. This suggests that MP intake for PAR ewes was likely around 330 g per day during lactation. Whilst a similar level of calculated MP intake significantly reduced worm egg output in ewes fed rations based on xylose-treated soya bean meal compared to a low MP control, the same level of calculated MP intake failed to reduce worm egg output in ewes fed rations based on faba beans (Sakkas et al., 2012). Furthermore, the high level of feed intake achieved in our ewes would have resulted in a large weight of faeces, which all else being equal would reduce the FEC due to its expression

as number of eggs per gram and thus its sensitivity to faecal dilution (Houdijk, 2008). Indeed, the calculated total worm egg output of approximately one million eggs per day around day₂₈ of lactation was in agreement with that observed when MP supply was scarce, even though FEC were about four times greater than in our PAR ewes (Kidane et al., 2010). Taken together, the above would support the view that immunity to parasites was at least to some extent compromised in our PAR ewes, in contrast to what the low FEC we reported here would intuitively indicate.

Ruminants lose 2 to 12% of their gross energy intake in the form of CH₄ (Johnson and Johnson, 1995), which accounts for a significant proportion of GHG emissions from livestock production systems. In our data, observed CH₄ yield averaged 10.23 g/kg DMI, equivalent to 3.3% of gross energy, which is relatively low compared to CH₄ energy losses on lucerne reported elsewhere, e.g. 4.7% on lucerne hay fed *ad libitum* (Pinares-Patinõ et al., 2003), 5.1% on pelleted lucerne fed restrictedly (Pinares-Patinõ et al., 2013; Waghorn et al., 2002), 5.9% on silage fed *ad libitum* (Bouchard et al., 2013), or 6.6% on freshly-cut and fed *ad libitum* (Waghorn et al., 2002). Furthermore, it was also lower than the 6.3% loss of CH₄ energy and out of the range of 3.7% to 13.3% reported from 61 studies in housed and grazing sheep fed a large variety of forage-based rations (McBride et al., 2013). There may be several reasons why CH₄ yield observed in our study was low. The high level of feed intake achieved in our ewes was likely a major contributor, since it is well established that CH₄ yield decreases with increased feed intake (Johnson and Johnson, 1995; Jentsch et al., 2007), which reduces rumen retention time (AFRC, 1993), and thus reduces CH₄ emissions (Goopy et al., 2014). Intake

was likely high since lucerne was offered as dried pellets rather than as long forage. In agreement with this observation, it has been reported that CH₄ yield on grass nuts was ~35% less than on fresh grass or grass silage in lowland replacement ewes (Aubry et al., 2014). In addition, subject to variety, environment and harvest date (Pecetti et al., 2006), lucerne is known to contain variable amounts of saponins, which may not only mitigate methanogenesis (Jayanegara et al., 2010; Cieslak et al., 2013), but may also be anthelmintic (Ali et al., 2011), and it can therefore not be excluded that this may also have contributed to the relatively low FEC observed.

In our experiment, observed CO₂ yield averaged 690 g per kg DMI, which agrees with the earlier respiration and carbon balance measurements using sheep, fed restricted amounts of dried grass (Blaxter and Graham, 1955; Midwood et al., 1994). Tables 3 and 4 show that breath CO₂ could be considered proportionally the largest component of GHG production, yields and intensities measured. However, since breath CO₂ is an inevitable consequence of metabolism, CO₂ contributions to GWP calculations are those arising from fossil fuel use only (Cederberg et al., 2013). However, even if we include breath CO₂ in our GWP calculations, the relative effect of parasitism on GHG intensity remains 16%. Within the animal derived GHG contributions, enteric CH₄, manure N₂O and manure CH₄ comprise 67, 30 and 3%, respectively for CON and RES ewes and 64, 32 and 4% respectively for PAR ewes in the final calculated GWP per kg lamb BWG figures. However, the relative contribution of these GHG sources to the increase in calculated GHG intensity averaged at 47, 43 and 7%, respectively. Whilst this indicates that enteric CH₄ remains the largest contributor of GWP per kg lamb BWG, enteric

CH₄ and manure N₂O contributed almost equally to the parasitism-increased GHG intensity.

We identified that the impact of parasitism on observed and calculated yields per kg dOMI was more pronounced than per kg DMI (Table 3). Although this was the direct consequence of impact of parasitism on total tract OM digestibility (Table 2), it raises the question what would be the most appropriate unit to express GHG yield. It might be argued that most studies resort to yield per unit DMI since data on total tract OM digestibility are not generally available. Since it has been suggested that animals aim to optimise ME intake through their feeding behaviour (Tolkamp, 2010), and ME is directly proportional to the digestible organic matter component of the ration, dOMI is an excellent proxy in the absence of respiration data. Furthermore, variation in GHG production is better explained per unit dOMI rather than DMI, as the former better accounts for differences in diet quality (Muetzel, 2009). In this respect, it might be argued that the 23% relative impact of parasitism on GHG intensity on the basis of yield per kg dOMI is not dissimilar to the 16% increase on the basis of GHG intensity per kg lamb BWG. Hence, when OM digestibility data are available, expression of GHG yield per unit dOMI may be preferred over GHG yield per unit DMI, as it would also be more reflective of the most preferred expression, i.e. GHG per unit product (Hristov et al., 2013).

The functional unit in our study was one kg of lamb BW gain between birth and weaning, and we used this to estimate the level of milk production (Robinson et al., 1969). Although the aim of our work was not to develop a full life cycle assessment for ewe milk production, the averaged calculated GHG intensity of 1.13 ± 0.02 kg CO₂-eq per kg milk fitted in the wide range of

intensities reported for sheep milk production from earlier life cycle assessments, i.e. from 0.9-1.7 kg CO₂-eq per kg milk (Haas et al., 2001) but was lower than more recent estimates, i.e. 1.8-4.5 kg CO₂-eq per kg milk (Batalla et al., 2004) and 8.4 CO₂-eq per kg milk (Opio et al., 2013). This likely reflects the development in life cycle assessment methodologies over the last few years, and an increased understanding of GHG sources that require to be taken into account (Cederberg et al., 2013). In the context of controlling parasitism, this would need to include GHG emissions arising from the intervention.

Using the reported impacts of parasitism on FCR and assuming no impact on GHG yields, it can be calculated that lamb parasitism may increase GHG intensities to greater magnitudes than observed here in ewes, by up to 21% under housed conditions (Kyriazakis et al., 1994; Zaralis et al., 2008) and up to 139% under field conditions (Thamsborg and Agergaard, 2002). These figures could increase significantly if lamb parasitism does increase GHG yield, which has yet to be established. Thus, improvement of animal health has great potential to contribute to climate change mitigation strategies (Shields and Orme-Evans, 2015). Indeed, a 10% reduction in calculated GHG intensity has been estimated from suppressive anthelmintic treatments in continuously naturally infected, growing lambs (Keynon et al., 2013), though in agreement with our observations, reduced exposure can be expected to result in an even greater performance benefit (Coop et al., 1982) and thus reduction in GHG intensity. However, studies that have examined implications of improving animal health on GHG emission intensity are scarce, despite that improving feed efficiency has been recognised as a major driver to reduce

GHG emissions (Basarab et al., 2013), and connections among animal health and resource efficiency are obvious (Hristov et al., 2013). Perhaps as a consequence, improving animal health as a climate change mitigation option is not often considered amongst other more technical options, including reducing undernourishment (Eckard et al., 2010; Gerber et al., 2013). The framework developed here and the results for the specific hypothesis tested, may provide a tool and impetus for further studies in this field, including assessing different nutritional and challenge environments. Furthermore, they may assist to ensure that improvement of animal health is an increasingly recognised and integrated component of efforts to reduce the environmental footprint of animal production systems.

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References

590 AFRC. 1993. Energy and protein requirements of ruminants. An Advisory
 591 Manual Prepared by the AFRC Technical Committee on Responses to
 592 Nutrients, CAB International, Wallingford.

593 Akinbamijo, O.O., Lahlou-Kassi, I.A., Tembely, S., 1994. Fascioliasis and
 594 nutrient metabolism in pregnant and non-pregnant sheep. In: Lebbie
 595 S.H.B., Kagwini E. (Eds.), Small Ruminant Research and Development
 596 in Africa. Proceedings of the 3rd Biennial Conference of the African Small
 597 Ruminant Research Network. ILRI, Nairobi.
 598 (<http://www.fao.org/wairdocs/ilri/x5473b/x5473b15.htm>)

599 Ali, N., Shah, S.W.A., Shah, I., Ahmed, G., Ghias, M., Khan, I. 2011. Cytotoxic
 600 and anthelmintic potential of crude saponins isolated from *Achillea*
 601 *Wilhelmsii* C. Koch and *Teucrium Stocksianum* boiss. BMC
 602 Complement. Altern. Med. 11, 106.

603 Aubry, A., Annett, R., Yan, T., 2014. Effects of breed and forage types on
 604 methane emission factors for lowland replacement ewes aged between
 605 8 and 19 months. Adv. Anim. Biosci. 5, 25.

606 Basarab, J.A., Beauchemin, K.A., Baron, V.S., Ominski, K.H., Guan, L.L.,
 607 Miller, S.P., Crowley, J.J., 2013. Reducing GHG emissions through
 608 genetic improvement for feed efficiency: effects on economically
 609 important traits and enteric methane production. Animal 7 (S2), 303–
 610 315.

611 Batalla, I., Pinto, M., Unamunzaga, O., Besga G., del Hierro, Ó., 2004.
 612 Integrating social and economic criteria in the carbon footprint analysis in
 613 sheep dairy farms. In: Schenck, R., Huizenga, D. (Eds.), Proceedings of

614 the 9th International Conference on Life Cycle Assessment in the Agri-
 615 Food Sector. ACLCA, Vashon, pp. 88–95.

616 Blaxter, K. L., Graham, N.McC., 1955. Plane of nutrition and starch
 617 equivalents. J. Agric. Sci. 46, 292–306.

618 Bouchard, K., Wittenberg, K.M., Legesse, G., Krause, D.O., Khafipour, E.,
 619 Buckley, K.E., Ominski, K.H., 2013. Comparison of feed intake, body
 620 weight gain, enteric methane emission and relative abundance of rumen
 621 microbes in steers fed sainfoin and lucerne silages under western
 622 Canadian conditions. Grass Forage Sci. 70, 116–129.

623 Cederberg, C., Henriksson, M., Berglund, M., An LCA researcher's wish list –
 624 data and emission models needed to improve LCA studies of animal
 625 production. Anim. 7 (S2), 212–219.

626 Christie, M., Jackson, F., 1982. Specific identification of strongyle eggs in
 627 small samples of sheep faeces. Res. Vet. Sci. 32, 113–117.

628 Cieslak, A., Szumacher-Strabel, M., Stochmal, A., Oleszek, W., 2013. Plant
 629 components with specific activities against rumen methanogens. Anim. 7
 630 (S2), 253–265.

631 Coop, R.L., Kyriazakis, I., 1999. Nutrition-parasite interaction. Vet. Parasitol.
 632 84, 187–204.

633 Coop, R.L., Sykes, A.R., Angus, K.W., 1982. The effect of three levels of
 634 intake of *Ostertagia circumcincta* larvae on growth rate, food intake and
 635 body composition of growing lambs. J. Agricul. Sci. 98, 247–255.

636 Eckard, R.J., Grainger, C., de Klein, C.A.M., 2010. Options for the abatement
 637 of methane and nitrous oxide from ruminant production: A review. Livest.
 638 Sci. 130, 47–56.

639 Gallego, A., Hospido, A., Moreira, M.T., Feijoo, G., 2011. Environmental
640 assessment of dehydrated alfalfa production in Spain. *Resour. Conserv.*
641 *Recy.* 55, 1005–1012.

642 Gerber, P.J., Hristov, A.N., Henderson, B., Makkar, H., Oh, J., Lee, C.,
643 Meinen, R., Montes, F., Ott, T., Firkins, J., Rotz, A., Dell, C., Adesogan,
644 A.T., Yang, W.Z., Tricarico, J.M., Kebreab, E., Waghorn, G., Dijkstra, J.,
645 Oosting, S. 2013. Technical options for the mitigation of direct methane
646 and nitrous oxide emissions from livestock: a review. *Anim.* 7 (S2), 220-
647 234.

648 Goopy, J.P., Donaldson, A., Hegarty, R.S., Vercoe, P.E., Haynes, F., Barnett,
649 M., Oddy, V.H., 2014. Low-methane yield sheep have smaller rumens
650 and shorter rumen retention time. *Br. J. Nutr.* 111, 578-585.

651 Gorniak, T., Meyer, U., Südekum, K.-H., Dänicke, S., 2014. Effect of ambient
652 temperature on nutrient digestibility and nitrogen balance in sheep fed
653 brown-midrib maize silage. *Arch. Anim. Nutr.* 68, 336–344.

654 Haas, G., Wetterich, F., Köpke, U., 2001. Comparing intensive, extensified
655 and organic grassland farming in southern Germany by process life
656 cycle assessment. *Agricul. Ecosys. Environ.* 83, 43–53.

657 Hazzledine, M., 2014. Premier Atlas Ingredients Matrix. Premier Nutrition
658 Products LTD, Staffs, UK.

659 Houdijk, J.G.M., Kyriazakis, I., Jackson, F., Huntley, J.F. and Coop, R.L.,
660 2003. Is the allocation of metabolisable protein prioritised to milk
661 production rather than to immune functions in *Teladorsagia circumcincta*
662 infected lactating ewes? *Int. J. Parasitol.* 33, 327–338.

663 Houdijk, J.G.M., Jackson, F., Coop, R.L., Kyriazakis, I., 2006. Rapid
 664 improvement of immunity to *Teladorsagia circumcincta* is achieved
 665 through a reduction in the demand for protein in lactating ewes. *Int. J.*
 666 *Parasitol.* 36, 219–227.

667 Houdijk, J.G.M., Kyriazakis, I., Kidane, A., Athanasiadou, S., 2012.
 668 Manipulating small ruminant parasite epidemiology through the
 669 combination of nutritional strategies. *Vet. Parasitol.* 186, 38–50.

670 Houdijk, J.G.M., 2008. Influence of periparturient nutritional demand on
 671 resistance to parasites in livestock. *Par. Immunol.* 30, 113–121.

672 Hristov, A.N., Oh, J., Lee, C., Meinen, R., Montes, F., Ott, T., Firkins, J., Rotz,
 673 A., Dell, C., Adesogan, A., Yang, W., Tricarico, J., Kebreab, E.,
 674 Waghorn, G., Dijkstra, J., Oosting, S., 2013. Mitigation of greenhouse
 675 gas emissions in livestock production – A review of technical options for
 676 non-CO₂ emissions. In: Gerber, P.J., Henderson B., Makkar, H.P.S.,
 677 (Eds.), *FAO Animal Production and Health Paper No. 177*. FAO, Rome.

678 IPCC, 2006. *Guidelines for National Greenhouse Gas Inventories*.
 679 Intergovernmental panel on Climate Change.

680 Jayanegara, A., Goel, G., Makkar, H.P.S., Becker, K., 2010. Reduction in
 681 methane emissions from ruminants by plant secondary metabolites:
 682 effects of polyphenols and saponins. In: Odongo, N.E., Garcia, M.,
 683 Viljoen, G.J., (Eds.), *Sustainable Improvement of Animal Production and*
 684 *Health*. FAO, Rome, pp. 151–157.

685 Jentsch W., Schweigel M., Weissbach F., Scholze H., Pitroff W., Derno M.,
 686 2007. Methane production in cattle calculated by the nutrient
 687 composition of the diet. *Arch. Anim. Nutr.* 61, 10–19.

688 Johnson, K.A., Johnson, D.E., 1995. Methane emissions from cattle. J. Anim.
689 Sci. 73, 2483–2492.

690 Kenyon, F., Dick, J.M., Smith, R.I., Coulter, D.G., McBean, D., Skuce, P.J.,
691 2013. Reduction in greenhouse gas emissions associated with worm
692 control in lambs. Agric. 3, 271–284.

693 Kidane, A., Houdijk, J.G.M., Athanasiadou, S., Tolkamp, B.J., Kyriazakis, I.,
694 2010. Nutritional sensitivity of periparturient resistance to nematode
695 parasites in two breeds of sheep with different nutrient demands. Br. J.
696 Nutr. 104, 1477–1486.

697 Kidane, A., Houdijk, J.G.M., Tolkamp, B.J., Athanasiadou, S., Kyriazakis, I.,
698 2009. Consequences of infection pressure and protein nutrition on
699 periparturient resistance to *Teladorsagia circumcincta* and performance
700 in ewes. Vet. Parasitol. 165, 78–87.

701 Kimambo, A.E., Macrae, J., C., Dewey, P.J.S., 1988. The effect of daily
702 challenge with *Trichostrongylus colubriformis* larvae on the nutrition and
703 performance of immunologically-resistant sheep. Vet. Parasitol. 28,
704 205–212.

705 Kyriazakis, I., Oldham, J.D., Coop, R.L., Jackson, F., 1994. The effect of
706 subclinical intestinal nematode infection on the diet selection of growing
707 sheep. Br. J. Nutr. 72, 665–677.

708 Kyriazakis, I., Tolkamp, B.J., Hutchings, M.R., 1998. Towards a functional
709 explanation for the occurrence of anorexia during parasitic infections.
710 Anim. Behav. 56, 265–274.

711 Leyva, V., Henderson, A.E., Sykes, A.R., 1982. Effect of daily infection with
 712 *Ostertagia circumcincta* larvae on food intake, milk production and wool
 713 growth in sheep. J. Agric. Sci. 99, 249–259.

714 Lima, R., Díaz, R.F., Castro, A., Fievez, V., 2011. Digestibility, methane
 715 production and nitrogen balance in sheep fed ensiled or fresh mixtures
 716 of sorghum–soybean forage. Livest. Sci. 141, 36–46.

717 Lynch, G.P., Elsasser, T.H., Rumsey, T.S., Jackson Jr, C. and Douglass, L.W.
 718 1988. Nitrogen metabolism by lactating ewes and their lambs. J. Anim.
 719 Sci. 66, 3285–3294.

720 Maâmouri, O., Atti, N., Mahouachi, M. and Kraeim, K., 2011. Effect of grass
 721 nitrogen protection by *Acacia cyanophylla* on nitrogen balance and
 722 sheep milk production. In: Bouche, R., Derkimba, A., Casabianca, F.
 723 (Eds.), New trends for innovation in the Mediterranean animal
 724 production. Wageningen Academic Publishers, Wageningen, pp. 153–
 725 156.

726 Malik, B., Nicol A.M., van Houtert, M., 1999. Source of excess nitrogen affects
 727 nutrient partitioning in lactating ewes. Proc. New Zeal. Soc. Anim. Prod.
 728 59, 158–161.

729 McBride, J., Morrison, S.J., Yan, T., 2013. Review of enteric methane
 730 emissions of cattle and sheep fed diets relevant to UK farming
 731 conditions. Adv. Anim. Biosci. 4 (2), 299.

732 Midwood, A.J., Haggarty, P., McGaw, B.A., Mollison, G.S., Milne, E., Duncan,
 733 G.J., 1994. Validation in sheep of the doubly labelled water method for
 734 estimating CO₂ production. Am. J. Physiol. 266, R169–R179.

735 Ministry of Agriculture Fisheries and Food, 1992. Analysis of Agricultural
736 Materials, 2nd Ed. Her Majesty's Stationary Office, London.

737 Muetzel, S., 2009. Effect of level of intake on methane production per kg of
738 dry matter intake. MAF Technical Paper No: 2011/95.

739 Olthoff, J.C., Dickerson, G.E., Nienaber, J.A., 1989. Energy utilization in
740 mature ewes from seven breeds with diverse production potentials. J.
741 Anim. Sci. 67, 2550-2564.

742 Opio, C., Gerber, P., Mottet, A., Falcucci, A., Tempio, G., MacLeod, M.,
743 Vellinga, T., Henderson, B., Steinfeld, H., 2013. Greenhouse gas
744 emissions from ruminant supply chains. A global life cycle assessment.
745 FAO, Rome.

746 Pappas, A., 1977. Protein requirements of lactating Chios ewes. J. Anim. Sci.
747 44, 672–679.

748 Parkins, J.J., Holmes, P.H., Bremner, K.C., 1973. The pathophysiology of
749 ovine ostertagiasis: some nitrogen balance and digestibility studies. Res.
750 Vet. Sci. 14, 21–28.

751 Pecetti, L., Tava, A., Romani, M., De Benedetto, M.G., Corsi, P., 2006.
752 Variety and environment effects on the dynamics of saponins in lucerne
753 (*Medicago sativa* L.). Eur. J. Agron. 25, 187–192.

754 Pinares-Patinõ, C.S., Hickey, S.M., Young, E.A., Dodds, K.G., MacLean, S.,
755 Molano, G., Sandoval, E., Kjestrup, H., Harland, R., Hunt, C., Pickering
756 N.K., McEwan J.C. 2013. Heritability estimates of methane emissions
757 from sheep. Anim. 7(s2), 316–321.

758 Pinares-Patinõ, C.S., Ulyatt, M.J., Waghorn, G.C., Lassey, K.R., Barry, T.N.,
759 Holmes, C.W., Johnson, D.E. 2003. Methane emission by alpaca and

760 sheep fed on lucerne hay or grazed on pastures of perennial
 761 ryegrass/white clover or birdsfoot trefoil. J. Agric. Sci. 140, 215–226.

762 Robinson, J.J., Foster, W.H., Forbes, T.J., 1969. The estimation of the milk
 763 yield of a ewe from body weight data on the suckling lamb. J. Agric. Sci.
 764 Camb. 72, 103–107.

765 Rooke, J.A., Wallace, R.J., Duthie, C.-A., McKain, N., Motta de Souza, S.,
 766 Hyslop, J.J., Ross, D.W., Waterhouse, T., Roehe, R., 2014. Hydrogen
 767 and methane emissions from beef cattle and their rumen microbial
 768 community vary with diet, time after feeding and genotype. Br. J. Nutr.
 769 112, 398–407.

770 Russel, A.J.F., Doney, J.M., Gunn, R.G., 1969. Subjective assessment of
 771 body fat in live sheep. J. Agric. Sci. Camb. 72, 451–454.

772 Sakkas, P., Houdijk, J.G.M., Athanasiadou, S., Kyriazakis, I., 2012. Sensitivity
 773 of periparturient breakdown of immunity to parasites to dietary protein
 774 source. J. Anim. Sci. 90, 3954–3962.

775 Shields, S., Orme-Evans, G., 2015. The impacts of climate change mitigation
 776 strategies on animal welfare. Anim. 5, 361–394.

777 Sykes, A.R., Coop, R.L., 1977. Intake and utilization of food by growing sheep
 778 with abomasal damage caused by daily dosing with *Ostertagia*
 779 *circumcincta* larvae. J. Agric. Sci., Camb. 88, 671–677.

780 Sykes, A.R., 1994. Parasitism and production in farm animals. Anim. Prod.
 781 59, 155–172.

782 Thamsborg, S.M., Agergaard, N., 2002. Anorexia and food utilization in
 783 nematode infected lambs on pasture. Anim. Sci. 75, 303–313.

784 Thomas, P.C., Roberston, S., Chamberlain, D.G., Livingstone, R.M.,
 785 Garthwaite, P.H., Dewey, P.J.S., Smart, R., Whyte, C., 1988. Predicting
 786 the metabolizable energy content of compounded feeds for ruminants.
 787 In: Haresign, W., Cole, D.J.A., (Eds.), Recent Advances in Animal
 788 Nutrition. Butterworths, London, pp. 127–146.

789 Tolkamp, B.J., 2010. Efficiency of energy utilisation and voluntary feed intake
 790 in ruminants. *Anim.* 4, 1084–1092.

791 van Keulen, J., Young, B.A., 1977. Evaluation of the acid-insoluble ash as a
 792 natural marker in ruminant digestibility studies. *J. Anim. Sci.* 44, 282–
 793 287.

794 Waghorn, G.C., Tavendale, M.H., Woodfield, D.R., 2002. Methanogenesis
 795 from forages fed to sheep. *Proc. New Zeal. Grassl. Assoc.* 64, 167–171.

796 Zaralis, K., Tolkamp, B.J., Houdijk, J.G.M, Wylie, A.R.G., Kyriazakis, I. 2008.
 797 Food intake and plasma leptin concentrations during gastrointestinal
 798 parasitism in lambs of two breeds. *J. Anim. Sci.* 86, 1891–1903.

799 Zaralis, K., Tolkamp, B.J., Houdijk, J.G.M, Wylie, A.R.G., Kyriazakis, I., 2009.
 800 Consequences of protein supplementation on anorexia, expression of
 801 immunity and plasma leptin concentrations in parasitised ewes of two
 802 breeds. *Br. J. Nutr.* 101, 499–509.

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Legends to figures

Figure 1. Dry matter intake of twin-rearing ewes fed pelleted lucerne, either sham-infected and fed *ad libitum* (○), or trickle-infected with *Teladorsagia circumcincta* and fed *ad libitum* (●), or sham-infected and fed restrictedly during lactation at 80% of intake by ewes fed *ad libitum* (◇).

Figure 2. Hourly methane and carbon dioxide production of pairs of pelleted lucerne fed twin-rearing ewes at 5 wks into lactation, either sham-infected and fed *ad libitum* (○), or trickle-infected with *Teladorsagia circumcincta* and fed *ad libitum* (●), or sham-infected and fed restrictedly during lactation at 80% of intake by ewes fed *ad libitum* (◇). The arrow indicates time of feeding.

820 Table 1. Analysed composition of lucerne.

821

Analysis	
Dry matter (g/kg)	974
Neutral detergent fibre (g/kg DM)	448
Acid detergent fibre (g/kg DM)	362
Crude protein (6.25×N, g/kg DM)	163
Ash (g/kg DM)	103
Acid hydrolysis ether extract (AH-EE, g/kg DM)	19.4
Acid insoluble ash (g/kg DM)	13.7
<i>In vitro</i> organic matter digestibility (NCGD ¹ , %)	57.2
Gross energy (MJ/kg DM)	17.9
Digestible energy ² (MJ/kg DM)	10.2
Metabolizable energy ³ (MJ/kg DM)	8.3

822

823 ¹Neutral cellulose and gammanase digestibility

824 ²Calculated as ME/0.81 (AFRC, 1993)

825 ³Calculated from AH-EE and NCGD (Thomas et al., 1988).

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827 Table 2. Apparent feed digestibility and ewe performance during lactation of
 828 twin-rearing ewes fed pelleted lucerne, either sham-infected and fed *ad*
 829 *libitum* (CON), or trickle-infected with *Teladorsagia circumcincta* and fed *ad*
 830 *libitum* (PAR), or sham-infected and fed restrictedly during lactation at 80% of
 831 intake by ewes fed *ad libitum* (RES).

832

	Treatments ¹			s.e.d.
	CON	PAR	RES	
Apparent digestibility				
Dry matter (DM, %)	53.8 ^a	45.6 ^b	55.3 ^a	2.98
Organic matter (OM, %)	54.6 ^a	46.4 ^b	55.8 ^a	2.99
Crude protein (CP, %)	58.8 ^a	53.4 ^b	61.9 ^a	2.87
Performance				
DM intake (kg/day)	4.58 ^a	4.29 ^b	3.55 ^c	0.10
Digestible OM intake (kg/day)	2.24 ^a	1.78 ^b	1.77 ^b	0.13
Digestible CP intake (kg/day)	0.44 ^a	0.37 ^b	0.36 ^b	0.02
Ewe body weight gain (g/day)	-69 ^a	-162 ^b	-252 ^c	37
Litter body weight gain (g/day)	718 ^a	669 ^b	657 ^b	24
Estimated milk production (g/day)	3524 ^a	3223 ^b	3142 ^b	149

833

834 ¹Data within the same row with different superscripts differ (P<0.05).

835 Table 3. Observed and calculated greenhouse gas production and yield
836 during lactation of twin-rearing ewes fed pelleted lucerne, either sham-
837 infected and fed *ad libitum* (CON), or trickle-infected with *Teladorsagia*
838 *circumcincta* and fed *ad libitum* (PAR), or sham-infected and fed restrictedly
839 during lactation at 80% of intake by ewes fed *ad libitum* (RES).

	Treatments ¹			s.e.d.
	CON	PAR	RES	
Enteric methane (observed)				
CH ₄ production (g/day/ewe)	55 ^a	49 ^b	41 ^c	2.8
(CO ₂ -eq)	1375 ^a	1225 ^b	1025 ^c	70
CH ₄ yield (g/kg DMI)	10.6	10.3	10.3	0.56
(CO ₂ -eq)	265	258	258	14
CH ₄ yield (g/kg dOMI)	22.0 ^a	25.2 ^b	20.9 ^a	1.54
(CO ₂ -eq)	550 ^a	630 ^b	523 ^a	39
Manure methane (calculated)				
Volatile solids production (kg/day)	2.24 ^a	2.44 ^a	1.69 ^b	0.12
CH ₄ production (g/day/ewe)	2.86 ^a	3.10 ^a	2.15 ^b	0.14
(CO ₂ -eq)	72 ^a	78 ^a	54 ^b	4
CH ₄ yield (g/kg DMI)	0.56 ^a	0.65 ^b	0.54 ^a	0.03
(CO ₂ -eq)	14 ^a	16 ^b	14 ^a	1
CH ₄ yield (g/kg dOMI)	1.22 ^a	1.79 ^b	1.15 ^a	0.24
(CO ₂ -eq)	30 ^a	45 ^b	29 ^a	6
Manure nitrous oxide (calculated)				
N excretion (g/day)	91.6 ^a	89.6 ^a	68.7 ^b	2.77
N ₂ O production (g/day/ewe)	2.05 ^a	2.01 ^a	1.54 ^b	0.06
(CO ₂ -eq)	611	599	459	18
N ₂ O yield (g/kg DMI)	0.40 ^{ab}	0.42 ^a	0.39 ^b	0.01
(CO ₂ -eq)	119 ^{ab}	125 ^a	116 ^b	3
N ₂ O yield (g/kg dOMI)	0.85 ^a	1.12 ^b	0.81 ^a	0.12
(CO ₂ -eq)	253 ^a	334 ^b	241 ^a	36
Methane and nitrous oxide combined as CO ₂ -eq (calculated)				
GHG production (g/day/ewe)	2044 ^{a,x}	1909 ^{a,y}	1524 ^b	72
GHG yield (g/kg DMI)	399	397	386	14
GHG yield (g/kg dOMI)	844 ^a	1039 ^b	798 ^a	83
Breath carbon dioxide (observed)				
CO ₂ production (g/day/ewe)	3463 ^a	3241 ^a	2844 ^b	147
CO ₂ yield (g/kg DMI)	676	674	721	26
CO ₂ yield (g/kg dOMI)	1413 ^a	1664 ^b	1470 ^{ab}	111

840 ¹Data with different superscripts differ (^{a,b,c}P<0.05; ^{x,y}P<0.10).

Table 4. Effects of maternal parasitism and restricted feeding on time and dry matter intake (DMI) required to wean two lambs at 25 kg each and compensate for loss of body weight (BW), and its effect on calculated ewe global warming potential (GWP) of twin-rearing ewes fed pelleted lucerne, either sham-infected and fed *ad libitum* (CON), or trickle-infected with *Teladorsagia circumcincta* and fed *ad libitum* (PAR), or sham-infected and fed restrictedly during lactation at 80% of intake by ewes fed *ad libitum* (RES).

		Treatments ¹			s.e.d.
		CON	PAR	RES	
Calculated performance and manure production parameters					
Days to target (n)		57.1 ^a	61.8 ^b	62.7 ^b	2.39
Total BW lost (kg)		3.4 ^a	10.0 ^b	14.4 ^c	1.85
DMI to target (kg)		253 ^a	257 ^a	214 ^b	9.5
DMI to compensate BW loss (kg)		16 ^a	60 ^b	68 ^b	9.5
Total DMI (kg)		270 ^a	316 ^b	283 ^a	11.6
Feed conversion ratio ²		6.7 ^a	7.8 ^b	7.0 ^a	0.31
Manure volatile solids (kg)		115 ^a	151 ^b	118 ^a	10.2
Manure N output (kg)		4.75 ^a	6.11 ^b	5.10 ^a	0.28
Calculated GWP (kg CO ₂ -eq/kg lamb body weight gain)					
Enteric	CH ₄	1.79 ^a	2.00 ^b	1.80 ^a	0.10
Manure	N ₂ O	0.80 ^a	1.00 ^b	0.87 ^a	0.05
	CH ₄	0.09 ^a	0.12 ^b	0.10 ^a	0.01
Feed	CO ₂ -eq	2.41 ^a	2.79 ^b	2.51 ^a	0.11
Total GWP		5.09 ^a	5.91 ^b	5.28 ^a	0.21
Calculated breath CO ₂ (g/kg lamb body weight gain)					
		4.52 ^a	5.21 ^b	4.96 ^b	0.19

¹Data within the same row with different superscripts differ (^{a,b,c}P<0.05).

²Feed conversion ratio is calculated as total DMI divided over total lamb BW gain.